

## Influence of concentration and mode of application of different growth regulators on dendrobium hybrid *Thongchai Gold*

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### ABSTRACT

An experiment was conducted to determine the influence of concentration and mode of application of different growth regulators on growth and yield of *Dendrobium* hybrid “Thongchai Gold” in subtropical cool climate zone. Various concentrations of GA<sub>3</sub> and BA were applied as foliar spray and drenching mode. GA<sub>3</sub> 100 ppm in combination with BA 100 ppm followed by GA<sub>3</sub> 200 ppm increased the plant height (31.63 cm), number of shoots per plant (5.17), cane girth (1.71 cm), number of leaves per cane (8.21), internodal length (3.19 cm), spike girth (0.45 cm), number of flowers per spike (12.63) and number of spikes per cane (2.80). The spike length (29.38 cm) was noticed highest in GA<sub>3</sub> 50 ppm and rachis length (19.66 cm) was observed highest in GA<sub>3</sub> 200 ppm. Growth regulators and mode of applications also influenced nutrient content (NPK) of plants. Among the mode of applications, media drenching with the different growth regulator treatment was found to be best for the *Dendrobium* orchids.

**Keywords:** Drenching, foliar spray, growth regulators, yield

*Dendrobium*s are tropical orchids requiring warm, humid and shady conditions and the second largest genus of the orchid family with more than 1500 species. *Dendrobium* production has steadily increased with the influx of new farms being developed and with established farms that have increased in acreage. In India commercially grown in the states of Kerala, Tamil Nadu, Karnataka and Maharashtra, these orchids are easy to cultivate and produce abundant, colourful flowers. Plant growth regulators have been used successfully to modify flowering of many orchid species. Apical dominance and auxins suppress flowering of *Aranda* (Goh, 1985). The response of BA depends on the hybrids and successfully induced flowering in *Aranthera*, *Aranda*, *Holtumara*, *Mokara*, *Dendrobium Lousiae* 'Dark', *Phalaenopsis*, *Dendrobium 'Nodoka'*, and *Oncidium* (Goh, 1985; Hew and Clifford,1993). Gibberellins transformed the normal vegetative shoot apex to a terminal inflorescence of *Aranda* 'Deborah' and induced flowering of *Bletilla striata*, *Cymbidium* and *Cattleya* hybrids (Goh *et al.*, 1982; Goh, 1985). Addition of GA<sub>3</sub> with cytokinins enhanced the flowering effect and reduced flower deformity of *Phalaenopsis* and *Dendrobium* (Chen *et al.*, 1997; Sakai *et al.*, 2000). Injection of 100 mm of BA and 10mm GA<sub>3</sub> into *Dendrobium* 'Jaquelyn Thomas Uniwai Princess' pseudobulbs promoted off season flowering and increased the flower size and number of inflorescences per stem in Hawaii (Sakai *et al.*, 2000).

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Application of cytokinin also induced flowering in *Dendrobium*. Earlier evidence showed that some *Dendrobium* species can be induced to flower by either cytokinin treatment or following an expose to low temperature treatments. More over flowering of *Dendrobium* depends on genetic makeup of species, hybrids and climate condition of particular area where it is available. Application of exogenous growth regulators to hybrids for enhancement of production is limited. In commercial orchid cut flower production, the production should have coincidence with market demand. Induction of flower and seasonality of flowering are the two major factors that govern pricing and marketability of orchid cut flowers. Keeping the above condition in view, the present study was undertaken to study the influence of different growth regulators (BA and GA<sub>3</sub>) at different concentration and mode of application on *Dendrobium* hybrid “Thongchai Gold” for flower production under sub tropical cool climate of Sikkim.

### MATERIALS AND METHODS

An experiment was conducted at National Research Centre for Orchids, Pakyong, Sikkim during the period of 2008-2012. Tissue cultured plants of uniform size of *Dendrobium* hybrid “Thongchai Gold” were grown in partially modified green house. Plants were grown in 15 cm perforated pot with a growing medium of broken bricks (2 to 3cm size) and coconut husk (1.5 to 3cm size) at a ratio of 1:1 Initial two years plants were grown for acclimatization with the sub tropical cool climate where average maximum

air temperature ranged between 25°C to 30°C from April to September. The air temperature goes down slowly from end of October to February. During winter season minimum air temperature were recorded between 5°C to 10°C. New shoots of plant generally came up in the month of May to September and growth seized in winter month. Seven growth regulator treatments and two mode of application such as foliar spray and media drenching were followed at morning hours only. Plants were sprayed and drenched thoroughly with growth regulators of the consequent treatments such as distilled water as control, GA<sub>3</sub> 50 ppm, GA<sub>3</sub> 100ppm, GA<sub>3</sub> 200 ppm, BA 50 ppm, BA 100 ppm and GA<sub>3</sub> 100 ppm +BA 100 ppm. BA and GA<sub>3</sub> were dissolved in 95% ethanol at the concentrations required for each treatments and distilled water was added to make the final concentration of 10% ethanol. In case of drenching, complete saturation of growing medium with growth regulator solutions, excess solutions that passed through the pot holes were collected in tray and reused several times. In combine application, BA 100 ppm spray or drench was given 10 days after GA<sub>3</sub> spray or drenching of solution. Foliar application and drenching of chemicals were given at twice a year during the months of April and July (2011 and 2012). The water soluble fertilizer (NPK 20:20:20) was applied @ 1g/l as foliar spray at weekly intervals initially for two years and NPK (10:20:20) @1g/l from third year onward for boosting vegetative and flowering growth respectively.

The observations on plant height (cm), number of shoots per plant, cane girth (cm), number of leaves per cane, inter-nodal length (cm), spike length (cm), rachis length (cm), spike girth (cm), number of flowers per spike and number of spikes per cane were recorded periodically at regular intervals. The experiment was carried out in a factorial completely randomized design and replicated thrice. Total N was determined by micro Kjeldahl method (Bremner, 1965), total P was determined by Vanodomolybdo-phosphoric yellow colour method (Jackson, 1973) and total K was determined by flame photometer method (Jackson, 1973). The pooled data were subjected to statistical analysis by using the software SAS, following the user's guide SAS/STAT (1990).

## **RESULTS AND DISCUSSION**

The growth of Dendrobium hybrid "Thongchai Gold" was significantly influenced by different concentrations of growth regulators and mode of application (Table 1). The plant height was recorded highest in the treatment combination of GA<sub>3</sub> 100 ppm

+ BA 100 ppm (31.63 cm) and among the mode of applications, drenching (29.44 cm) with the above growth regulator treatments was found to be best and in case of interaction, significant influence was noticed. According to Karaguzel *et al.* (2004) soil drenches were effective as a method of plant growth regulator application compared to foliar sprays. Flowering of Dendrobium is directly correlated with the cane or pseudobulb growth and development. So, the effect of the different concentrations of plant growth regulators and mode of applications on number of leaves per cane, cane girth, number of canes and intermodal length of the cane were studied. Application of GA<sub>3</sub> 100 ppm + BA 100 ppm consistently increased the shoots production (5.17) and followed by GA<sub>3</sub> 200 ppm (4.93). Among the mode of applications, there was no significant difference noticed, however significant influence was recorded in case of interaction of growth regulator and mode of applications. Application of BA 100 ppm in combination with GA<sub>3</sub> 100 ppm and BA 100 ppm alone increased the girth of the cane (1.71 and 1.60 cm respectively) and among the mode of applications, drenching recorded the highest cane girth (1.51 cm) as compared to foliar spraying. The interaction effect shows the significant difference between the mode of application and growth regulators. The treatment combination of GA<sub>3</sub> 100 ppm + BA 100 ppm increased the number of leaves per cane (8.21) followed by GA<sub>3</sub> 200 ppm (8.00) and among the mode, drenching of growth regulators (7.38). Application of GA<sub>3</sub> 200 ppm improved internodal length (3.22cm). The significant influence was recorded in case of interaction of growth regulator and mode of applications with the above growth regulator combinations. Reduction of internodal length was observed when BA was applied alone. However, the detrimental effect was reduced and inter nodal length was evident highest in plants which received GA<sub>3</sub> 100 ppm in combination with BA 100 ppm followed by GA<sub>3</sub> 200 ppm alone. Application of BA treatments promoted vegetative growth instead of flowering and this effect was alleviated by the addition of GA<sub>3</sub>. In Phalaenopsis cv. 'Leda', flower primordial elongation promoted by GA<sub>3</sub> is fully prevented by BA treatments (Chen *et al.*, 1997).

The perusal of table 2 shows that application of GA<sub>3</sub> 50 ppm followed by GA<sub>3</sub> 100 ppm + BA 100 ppm increased the spike length (29.38 cm and 28.81 cm respectively) and there is no significant influence was observed among the mode of applications, however, remarkable influence was recorded in case of

interaction effect between the growth regulators and mode of applications. Sugar may be a component of the floral stimulus and its production in the apical meristem could be enhanced by GA<sub>3</sub> treatment. Chen *et al.* (1994) stated that the sucrose syntheses may be relatively important in the degradation of sucrose in GA<sub>3</sub> treated plants. The findings with GA<sub>3</sub> would seem to support nutrient diversion as a mode of action by which exogenously applied GA<sub>3</sub> could promote flowering in *Phalaenopsis* (Sachs *et al.*, 1979). Our findings with GA<sub>3</sub> would seem to support the evidence by many workers (Chen *et al.*, 1994; Chen *et al.*, 1997; Sakai *et al.*, 2000; Matsumoto, 2006). GA<sub>3</sub> stimulate the transport of assimilate through the phloem, or increase sucrose uptake, or act on metabolism of sucrose and its metabolic products (Morris and Arthur, 1984). Gibberellins are biochemically described as tetra cyclic diterpene acids and are associated with

flowering induction in several species. When exogenously applied, these plant growth regulators lead to petal growth and flowering induction in long-day plants under conditions of short days. *Philodendron* cv. Black Cardinal plants were induced to flower under non-inductive conditions through application of GA<sub>3</sub> at 125, 250, 500 and 1,000 mg L<sup>-1</sup>, increasing flowering percentage and inflorescence number per plant with increasing concentrations (Chen *et al.*, 2003). In flower cultivation, other species have their flowering induced by GA<sub>3</sub> applied via pulverization, such as *Dieffenbachia* (Henny, 1980), *Zantedeschia* (Corr and Widmer, 1987) and *Anthurium* (Henny and Hamilton, 1992). Chen *et al.* (1997) reported that flowering can be induced in *Phalaenopsis* cv. Leda with the application of GA<sub>3</sub> under conditions of high temperatures, non-inductive to flowering.

**Table 1: Influence of concentration and mode of application of growth regulators on growth of *Dendrobium* hybrid *Thongchai gold***

| Growth regulators                   | Plant height (cm) |       |       | Number of shoots plant <sup>-1</sup> |      |      | Cane girth (cm) |      |      | Number of leaves cane <sup>-1</sup> |      |      | Internodal length (cm) |      |      |
|-------------------------------------|-------------------|-------|-------|--------------------------------------|------|------|-----------------|------|------|-------------------------------------|------|------|------------------------|------|------|
|                                     | S                 | D     | Mean  | S                                    | D    | Mean | S               | D    | Mean | S                                   | D    | Mean | S                      | D    | Mean |
| Control                             | 24.67             | 25.35 | 25.01 | 3.26                                 | 3.54 | 3.4  | 1.13            | 1.25 | 1.19 | 5.48                                | 6.50 | 5.99 | 2.50                   | 2.50 | 2.50 |
| GA <sub>3</sub> 50 ppm              | 28.20             | 29.07 | 28.64 | 3.76                                 | 3.94 | 3.85 | 1.33            | 1.37 | 1.35 | 6.49                                | 6.63 | 6.56 | 2.59                   | 2.88 | 2.74 |
| GA <sub>3</sub> 100 ppm             | 28.70             | 30.60 | 29.65 | 4.45                                 | 4.11 | 4.28 | 1.38            | 1.45 | 1.42 | 6.86                                | 6.85 | 6.85 | 2.94                   | 3.09 | 3.02 |
| GA <sub>3</sub> 200 ppm             | 30.81             | 31.37 | 31.09 | 4.85                                 | 5.01 | 4.93 | 1.45            | 1.57 | 1.51 | 7.67                                | 8.33 | 8.00 | 3.18                   | 3.26 | 3.22 |
| BA 50 ppm                           | 25.80             | 27.59 | 26.70 | 4.10                                 | 4.29 | 4.20 | 1.40            | 1.48 | 1.44 | 6.79                                | 7.19 | 6.99 | 2.67                   | 2.83 | 2.75 |
| BA 100 ppm                          | 28.18             | 29.75 | 28.97 | 4.38                                 | 4.58 | 4.48 | 1.54            | 1.66 | 1.60 | 7.05                                | 7.72 | 7.39 | 2.87                   | 2.99 | 2.93 |
| GA <sub>3</sub> 100 ppm +BA 100 ppm | 30.90             | 32.36 | 31.63 | 4.96                                 | 5.38 | 5.17 | 1.63            | 1.79 | 1.71 | 7.99                                | 8.43 | 8.21 | 3.20                   | 3.17 | 3.19 |
| <b>Mean</b>                         | 28.18             | 29.44 |       | 4.25                                 | 4.41 |      | 1.41            | 1.51 |      | 6.90                                | 7.38 |      | 2.85                   | 2.96 |      |
| <b>SEm(±)</b>                       |                   |       |       |                                      |      |      |                 |      |      |                                     |      |      |                        |      |      |
| <b>S</b>                            | <b>0.46</b>       |       |       | <b>0.20</b>                          |      |      | <b>0.05</b>     |      |      | <b>0.19</b>                         |      |      | <b>0.07</b>            |      |      |
| <b>D</b>                            | <b>0.40</b>       |       |       | <b>0.14</b>                          |      |      | <b>0.04</b>     |      |      | <b>0.15</b>                         |      |      | <b>0.05</b>            |      |      |
| <b>S×D</b>                          | <b>0.61</b>       |       |       | <b>0.30</b>                          |      |      | <b>0.08</b>     |      |      | <b>0.25</b>                         |      |      | <b>0.11</b>            |      |      |
| <b>LSD(0.05)</b>                    |                   |       |       |                                      |      |      |                 |      |      |                                     |      |      |                        |      |      |
| <b>S</b>                            | <b>1.30</b>       |       |       | <b>0.57</b>                          |      |      | <b>0.15</b>     |      |      | <b>0.54</b>                         |      |      | <b>0.21</b>            |      |      |
| <b>D</b>                            | <b>1.13</b>       |       |       | <b>NS</b>                            |      |      | <b>0.10</b>     |      |      | <b>0.41</b>                         |      |      | <b>NS</b>              |      |      |
| <b>S×D</b>                          | <b>1.72</b>       |       |       | <b>0.83</b>                          |      |      | <b>0.22</b>     |      |      | <b>0.72</b>                         |      |      | <b>0.30</b>            |      |      |

**Table 2: Influence of concentration and mode of application of growth regulators on flowering parameters of Dendrobium hybrid *Thongchai gold***

| Growth regulators                   | Spike length (cm) |       |       | Rachis length (cm) |       |       | Spike girth (cm) |      |      | Number of flowers/spike |       |       | Number of spikes cane <sup>-1</sup> |      |      |
|-------------------------------------|-------------------|-------|-------|--------------------|-------|-------|------------------|------|------|-------------------------|-------|-------|-------------------------------------|------|------|
|                                     | S                 | D     | Mean  | S                  | D     | Mean  | S                | D    | Mean | S                       | D     | Mean  | S                                   | D    | Mean |
| Control                             | 22.24             | 22.89 | 22.57 | 15.26              | 15.43 | 15.35 | 0.36             | 0.38 | 0.37 | 9.06                    | 9.24  | 9.15  | 1.33                                | 1.58 | 1.45 |
| GA <sub>3</sub> 50 ppm              | 29.04             | 29.71 | 29.38 | 16.80              | 16.71 | 16.76 | 0.40             | 0.40 | 0.40 | 9.90                    | 10.34 | 10.12 | 1.63                                | 1.74 | 1.69 |
| GA <sub>3</sub> 100 ppm             | 27.79             | 28.43 | 28.11 | 18.05              | 18.19 | 18.12 | 0.42             | 0.42 | 0.42 | 10.68                   | 11.74 | 11.21 | 1.86                                | 2.10 | 1.98 |
| GA <sub>3</sub> 200 ppm             | 28.62             | 28.80 | 28.71 | 19.25              | 20.06 | 19.66 | 0.43             | 0.44 | 0.44 | 12.20                   | 12.99 | 12.60 | 2.21                                | 2.52 | 2.37 |
| BA 50 ppm                           | 28.55             | 26.45 | 27.50 | 15.53              | 16.71 | 16.12 | 0.40             | 0.42 | 0.41 | 10.43                   | 10.78 | 10.61 | 1.60                                | 1.28 | 1.44 |
| BA 100 ppm                          | 25.79             | 27.46 | 26.63 | 16.44              | 18.08 | 17.26 | 0.42             | 0.43 | 0.43 | 11.15                   | 12.34 | 11.75 | 1.52                                | 1.24 | 1.38 |
| GA <sub>3</sub> 100 ppm +BA 100 ppm | 28.01             | 29.61 | 28.81 | 18.64              | 19.99 | 19.32 | 0.44             | 0.46 | 0.45 | 12.03                   | 13.23 | 12.63 | 2.54                                | 3.05 | 2.80 |
| <b>Mean</b>                         | 27.16             | 27.76 |       | 17.14              | 17.88 |       | 0.41             | 0.42 |      | 10.80                   | 11.52 |       | 1.94                                | 2.24 |      |
| <b>SEm(±)</b>                       |                   |       |       |                    |       |       |                  |      |      |                         |       |       |                                     |      |      |
| <b>S</b>                            | <b>0.53</b>       |       |       | <b>0.51</b>        |       |       | <b>0.01</b>      |      |      | <b>0.28</b>             |       |       | <b>0.11</b>                         |      |      |
| <b>D</b>                            | <b>0.44</b>       |       |       | <b>0.35</b>        |       |       | <b>0.00</b>      |      |      | <b>0.23</b>             |       |       | <b>0.08</b>                         |      |      |
| <b>S×D</b>                          | <b>0.72</b>       |       |       | <b>0.73</b>        |       |       | <b>0.01</b>      |      |      | <b>0.37</b>             |       |       | <b>0.14</b>                         |      |      |
| <b>LSD(0.05)</b>                    |                   |       |       |                    |       |       |                  |      |      |                         |       |       |                                     |      |      |
| <b>S</b>                            | <b>1.48</b>       |       |       | <b>1.45</b>        |       |       | <b>0.0159</b>    |      |      | <b>0.80</b>             |       |       | <b>0.30</b>                         |      |      |
| <b>D</b>                            | <b>NS</b>         |       |       | <b>NS</b>          |       |       | <b>0.0124</b>    |      |      | <b>0.65</b>             |       |       | <b>0.23</b>                         |      |      |
| <b>S×D</b>                          | <b>2.04</b>       |       |       | <b>2.06</b>        |       |       | <b>0.0214</b>    |      |      | <b>1.06</b>             |       |       | <b>0.39</b>                         |      |      |

Rachis length (19.66 cm) was recorded highest with the application of GA<sub>3</sub> 200 ppm followed by GA<sub>3</sub> 100 ppm in combination with BA 100 ppm (19.32 cm) and the mode of applications also failed to show significant difference. Application of GA<sub>3</sub> 100 ppm in combination with BA 100 ppm resulted in highest spike girth of 0.45 cm and it was on par with GA<sub>3</sub> 200 ppm (0.44 cm). Among the mode of applications media drenching (0.42 cm) was found to be best and significant influence was recorded in case of interaction of growth regulator and mode of applications. Number of flowers per spike was recorded highest with the application of GA<sub>3</sub> 100 ppm in combination with BA 100 ppm (12.63) followed by GA<sub>3</sub> 200 ppm (12.60) and among the mode of applications media drenching (11.52) was found to be best. The significant difference was noticed between the interaction of different concentrations of growth regulators and mode of applications. Flowering is a process that plant changes from vegetative to reproductive phase, from leaf to flower (Boonyakiat, 2003; Taiz and Zeiger, 2006). This process is controlled by hormonal level (Ruamrungsri, 2004) and environmental condition (Hew and Yong, 1997). Application of GA<sub>3</sub> 100 ppm in combination with BA 100 ppm increased the number of spikes per cane

(2.80). BA application alone produced deformed spikes which adversely affected the appearance of the spike. However, this detrimental effect of BA was reduced when GA<sub>3</sub> was applied along with BA. The results are in line with the findings of Rajeevan and Swapna (2003). The results obtained by Higuchi and Sakai (1977) were partly in line with the results of present study. They observed reduced number of florets in Dendrobium with the application of BAP at 4000 mg/l but productivity was found to increase. According to Goh and Yang (1978) application of GA<sub>3</sub> when combined with BA improved flowering in Dendrobium hybrid 'Lady Hochoy'. In *Dendrobium 'Louisae'*, GA<sub>3</sub> alone had any effect on floral bud initiation, while GA<sub>3</sub> at 0.001M+BA 0.01M, floral buds were appeared earlier than BA alone, these floral buds were better development and reached 100% of flower (Goh, 1979).

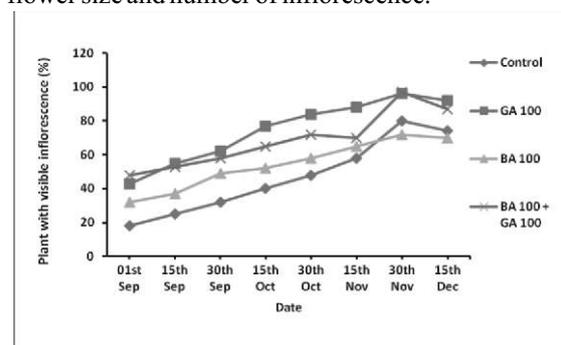
We observed deformed flower buds from plant treated with BA or GA<sub>3</sub>. Applications of GA<sub>3</sub> (Chen *et al.*, 1997) or BA (Sakai *et al.*, 2000) treatment to increase flowering often results in deformed flowers and usually require the combination of both for normal flower development. According to Blanchard and Runkle (2008) plants treated with BA<sub>1</sub>, BA<sub>2</sub>, or BA+GA within each treatment had flowers with

abnormal floral structures (for example, additional carpels, stamens, petals, or sepals); however, there were no trends observed among treatments. In *Phalaenopsis* 'Alice Girl,' BA1 applied 6 weeks after transfer to 23°C caused several plants to develop "crooked" inflorescences (for example, acute bend at 1 node). Kim *et al.* (2000) noted that 5–13% of *Doritaenopsis* plants treated with 400 mg L<sup>-1</sup> BA had malformed inflorescences at emergence and 3–7% of plants had blasted flower buds. In our study, application of BA on some orchid clones significantly reduced the width of open flowers by 1.2 cm or less. The decrease in flower number per inflorescence could be caused by competition among developing inflorescences for available assimilates. Additional research is required to elucidate the specific effects of BA on inflorescence development of these orchids.

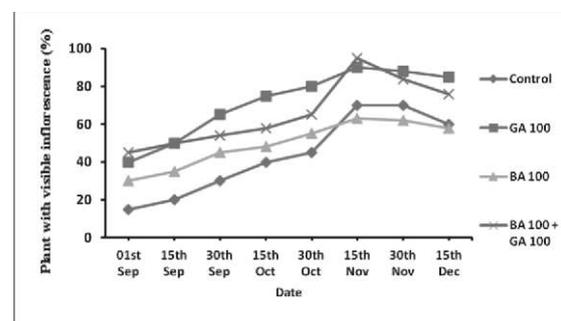
Treatment of *Phalaenopsis* flowering shoots with GA<sub>3</sub> at warmer temperatures increases levels of the active GA<sub>1</sub> required for the promotion of flower development to the same extent as levels found in flowering shoots grown at cool temperatures (Su *et al.*, 2001). We compared the percentage of plant with visible inflorescences in foliar spray and drenching of plant with growth regulators. It was observed that 15% plant produced inflorescence in control while 46% in the plant sprayed with BA 100 ppm GA<sub>3</sub> 100 ppm in 1<sup>st</sup> September followed by GA<sub>3</sub> 100 ppm. In all cases inflorescences increased as days progressed. The maximum inflorescence was recorded up to 15<sup>th</sup> November and decreased slowly thereafter (Fig.1). Over all; GA<sub>3</sub> 100 ppm enhanced the emergence of inflorescence. Same pattern of inflorescences development was observed in case of drenching, however maximum percentage of plants with inflorescence was recorded on 30<sup>th</sup> November and thereafter it decreased slowly. Like spraying, plants drenched with BA 100 ppm + GA<sub>3</sub> 100 ppm produced more number (97%) of inflorescence as compared to other treatments (Fig.2). The monthly production behavior was also noted during the experimentation period. It was recorded that the spraying of growth regulators influenced flower production for the period of September to December (Fig.3a,3b,3c,3d). There was an increasing trend of production at lower level of GA<sub>3</sub> application, however, the production fall at higher concentration (GA<sub>3</sub> 200ppm). The average production due to spraying of BA marginally increased at 100 ppm concentration.

The combine effect of GA 100ppm + BA 100ppm had improved the production more than 1.5 fold over control i.e. average 10.1 spike /pot/years in control and

16.55 in case of combined application of growth regulator. Same type of variation was observed in case of drenching of growth regulator (Fig.4a, 4b, 4c, 4d). The outstanding feature we observed that the production in drenched plant was increased more than the spraying irrespectively of growth regulator. The average production was around 2.5 fold more in combine application of GA<sub>3</sub> 100ppm + BA100ppm as compare to control. The lower production was caused in spraying invariably due to the plant of CAM in nature. During extreme winter the production was fallen drastically for which data was not presented. The cause is that most of the hybrids of *Dendrobium* are warm loving plant. Further, it might be in action of growth regulator during lower temperature. Matsumoto (2006) reported that application of BA treatments promoted vegetative growth instead of flowering and this effect was alleviated by addition of GA<sub>3</sub> in *Miltoniopsis* orchid hybrids Bert Field 'Eleen' and Rouge 'Akatsuka' which confirm our findings. In an experiment, Sakai *et al.* (2000) injected different concentration of BA and GA<sub>3</sub> and recorded 100mM of BA and 10mM GA<sub>3</sub> into the *Dendrobium* Jaquelyn Thomas 'Uniwai Princess' pseudobulb increased the flower size and number of inflorescence.



**Fig. 1: Percentage of plants with visible inflorescences as influenced by spraying of growth regulators**



**Fig. 2: Percentage of plants with visible inflorescences as influenced by drenching of growth regulators**

The table 3 depicts that the NPK content in leaves of Dendrobium hybrid Thongchai Gold was significantly influenced by the different concentrations of growth regulators, mode of applications and the interaction of both. Application of GA<sub>3</sub> 100 ppm in combination with BA 100 ppm increased the N (2.99), P (0.36) and K (3.35) content of leaves. Further, the mode of application as well as interaction with growth regulators also influenced the nutrients content. This might be due to increased nutrient content which would have contributed to the higher relative growth rate of the plants. Increased nutrient status in different plant parts are due to accumulation of carbohydrates, which may take place

gradually with the advancement of crop growth (Patil and Janawade, 1999).

For the foregoing discussion it can be concluded that the spike production, length of spike, rachis length and intermodal length are the commercially important characters in orchids. Considering these factors GA<sub>3</sub> 100 ppm combined with BA 100 ppm and GA<sub>3</sub> 200 ppm alone can be suggested as an ideal growth regulator combination for Dendrobium orchids at Sikkim condition. Our result shows that combination of both the growth regulators can be recommended to enhance the productivity without adversely affecting the quality of the spikes, however; further

**Table 3: Influence of concentration and mode of application growth regulators on NPK content of Dendrobium hybrid Thongchai gold**

| Growth regulators                   | Nitrogen    |      |      | Phosphorus  |      |      | Potassium   |      |      |
|-------------------------------------|-------------|------|------|-------------|------|------|-------------|------|------|
|                                     | S           | D    | Mean | S           | D    | Mean | S           | D    | Mean |
| Control                             | 2.73        | 2.78 | 2.76 | 0.26        | 0.27 | 0.27 | 3.07        | 3.10 | 3.09 |
| GA <sub>3</sub> 50 ppm              | 2.80        | 2.87 | 2.84 | 0.29        | 0.29 | 0.29 | 3.11        | 3.15 | 3.13 |
| GA <sub>3</sub> 100 ppm             | 2.83        | 3.02 | 2.93 | 0.30        | 0.31 | 0.31 | 3.13        | 3.20 | 3.17 |
| GA <sub>3</sub> 200 ppm             | 2.90        | 3.08 | 2.99 | 0.31        | 0.33 | 0.32 | 3.17        | 3.25 | 3.21 |
| BA 50 ppm                           | 2.67        | 2.83 | 2.75 | 0.30        | 0.31 | 0.31 | 3.20        | 3.26 | 3.23 |
| BA 100 ppm                          | 2.78        | 2.90 | 2.75 | 0.32        | 0.32 | 0.32 | 3.23        | 3.30 | 3.27 |
| GA <sub>3</sub> 100 ppm +BA 100 ppm | 2.87        | 3.10 | 2.99 | 0.35        | 0.36 | 0.36 | 3.30        | 3.40 | 3.35 |
| <b>Mean</b>                         | 2.80        | 2.94 |      | 0.34        | 0.31 |      | 3.17        | 3.24 |      |
| <b>SEm(±)</b>                       |             |      |      |             |      |      |             |      |      |
| <b>S</b>                            | <b>0.05</b> |      |      | <b>0.01</b> |      |      | <b>0.03</b> |      |      |
| <b>D</b>                            | <b>0.03</b> |      |      | <b>0.01</b> |      |      | <b>0.02</b> |      |      |
| <b>S×D</b>                          | <b>0.07</b> |      |      | <b>0.01</b> |      |      | <b>0.04</b> |      |      |
| <b>LSD(0.05)</b>                    |             |      |      |             |      |      |             |      |      |
| <b>S</b>                            | <b>0.16</b> |      |      | <b>0.02</b> |      |      | <b>0.08</b> |      |      |
| <b>D</b>                            | <b>0.09</b> |      |      | <b>0.02</b> |      |      | <b>0.06</b> |      |      |
| <b>S×D</b>                          | <b>0.19</b> |      |      | <b>0.02</b> |      |      | <b>0.11</b> |      |      |

Note: S- Spraying, D- Drenching

investigation is required for time and period of application.

#### REFERENCES

Blanchard, M. G. and Runkle, E. S. 2008. Benzyl adenine promotes flowering in *Doritaenopsis* and *Phalaenopsis* Orchids. *J. Pl. Growth Regul.*, **27**:141-50.

Boonyakiat, D. 2003. *Plant Physiology*. Faculty of Agriculture, Chiang Mai University, Chiang Mai. Thailand.

Bremner, J. M. 1965. Total nitrogen. In. *Methods of Soil Analysis. Part 2: Madison Agron. J.*, 1149-78.

Chen, J., Henny, R. J., McConnell, D. B. and Caldwell, R. D. 2003. Gibberellic acid affects growth and flowering of *Philodendron* ('Black Cardinal'). *Pl. Growth Regul.*, **41**: 1-6.

Chen, W. S., Liu, H. Y., Liu, Z.H., Yang, L. and Chen, W. H. 1994. Gibberellin and temperature influence carbohydrate content and flowering in *Phalaenopsis*. *Physiol. Pl.*, **90**:391-95.

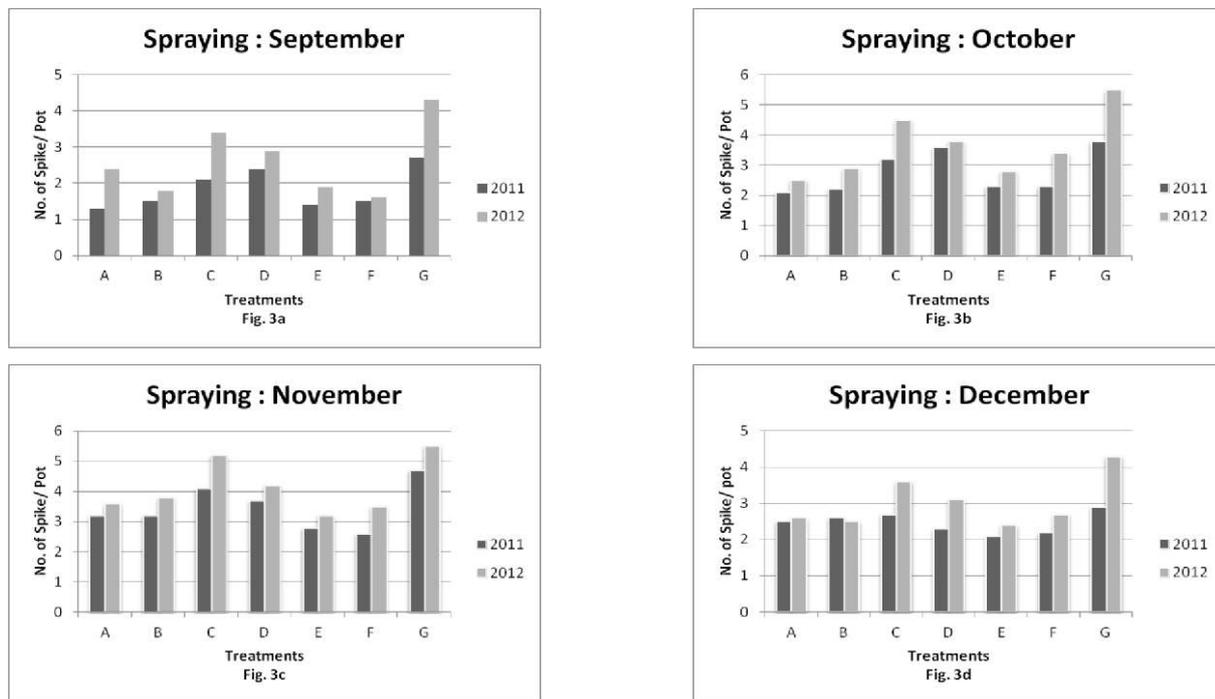


Fig. 3a, 3b, 3c, 3d showing the monthly production of spike<sup>-1</sup> pot by spraying of growth regulators. A-Control, B-GA<sub>3</sub> 50ppm, C- Ga<sub>3</sub> + 100ppm, D- Ga<sub>3</sub> + 200ppm, E-BA + 50ppm, F-BA + 100ppm, G-GA<sub>3</sub> + 100ppm BA100ppm.

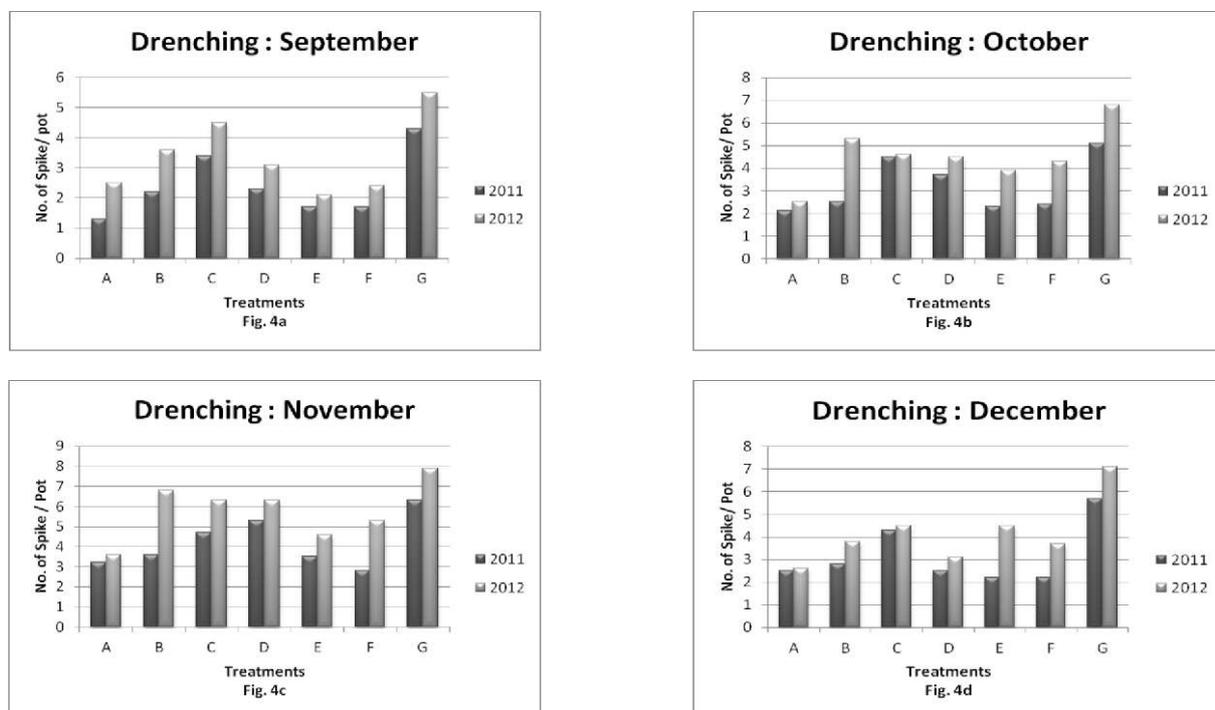


Fig. 4a, 4b, 4c, 4d showing the monthly production of spike<sup>-1</sup> pot by drenching of growth regulators. A-Control, B-GA<sub>3</sub>-50ppm, C- Ga<sub>3</sub> + 100ppm, D- Ga<sub>3</sub> + 200ppm, E-BA + 50ppm, F-BA + 100ppm, G-GA<sub>3</sub> + 100ppm BA100ppm.

- Chen, W. S., Chang, H. W., Chen, W. H. and Lin, Y. S. 1997. Gibberellic acid and cytokinin affect *Phalaenopsis* flower morphology at high temperature. *Hort. Sci.*, **32**: 1069-73.
- Corr, B. E. and Widmer, R. E. 1987. Gibberellic acid increases flower number in *Zantedeschia elliottiana* and *Z. rehmannii*. *Hort. Sci.*, **22**: 605-07.
- Goh, C. J. 1979. Hormonal regulation of flowering in a sympodial orchid hybrid *Dendrobium Louisae*. *New Phytol.*, **82**:375-80.
- Goh, C. J. 1985. Flowering in tropical orchids. *Proc. XI<sup>th</sup> World Orchid Conf.*, 17 May 1982, Florida. pp. 166-73.
- Goh, C. J. and Yang, A. L. 1978. Effects of growth regulators and decapitation on flowering of *Dendrobium* orchid hybrids. *Pl. Sci. Lett.*, **12**: 287-92.
- Goh, C. J., Strauss, M. S. and Arditti, J. 1982. Flower induction and physiology of orchid. In. *Orchid biology: Reviews and Perspectives* (Eds.) Cornell Univ. Press, Ithaca, New York, pp. 214-41.
- Henny, R. J. 1980. Gibberellic acid (GA<sub>3</sub>) induces flowering in *Dieffenbachia maculata* 'Perfection'. *Hort.Sci.*, **15**:613.
- Henny, R. J. and Hamilton, R. L. 1992. Flowering of *Anthurium* following treatment with gibberellic acid. *Hort. Sci.*, **27**: 1328.
- Hew, C. S. and Clifford, P. E. 1993. Plant growth regulators and the orchid cut-flower industry. *Pl. Growth Regul.*, **13**:231-39.
- Hew, C. S. and Yong, J. W. H. 1997. *The Physiology of Tropical Orchids in Relation to the Industry*. World Scientific Pub. Company Pvt. Ltd., Singapore, pp. 168-97.
- Highuchi, H. and Sakai, K. 1977. The effect of N6 – Benzyladenine on the flowering of *Dendrobium* 'Nodoka'. *Res. Bull. Aichi Ken Agri. Res. Centre, B. Hort.*, **9**: 79-81.
- Jackson, M. L. 1973. *Soil Chemical Analysis*. Prentice Hall India Pvt. Ltd. New Delhi, pp.38-82.
- Karaguzel, O., Baktir, I., Cakamakci, S. and Ortacesme, V. 2004. Growth and flowering response of *Lupinus varius* L. to paclobutrazol. *Hort. Sci.*, **39**: 1659-63.
- Kim, T. J., Lee, C. H. and Paek, K. Y. 2000. Effects of growth regulators under low temperature environment on growth and flowering of *Doritaenopsis* 'Happy Valentine' during summer. *J. Korean Soc. Hort. Sci.*, **41**:101-04.
- Matsumoto, T. K. 2006. Gibberilic acid and Benzyladenine promote early flowering and vegetative growth of *Miltoniopsis* orchid hybrids. *Hort. Sci.*, **41**: 131-35.
- Morris, D. A. and Arthur, E. D. 1984. Invertase activity in sinks undergoing cell expansion. *Pl. Growth Regul.*, **2**: 327-37.
- Patil, V. S. and Janawade, A. D. 1999. Soil water plant atmosphere relationships. *Proc. Adv. Micro Irrig. Fertig.*, 21-30 June, 1999, Dharwad, Karnataka, pp. 9-32.
- Rajeevan, P. K. and Swapna, S. 2003. Regulation of flower yield and quality in *Dendrobium Sonia* 17. *J. Orchid Soc. India*, **17**: 17-26.
- Ruamrungsri, S. 2004. *The Plant Physiology of Flower and Ornamental Plant*. Faculty of Agriculture, Chiang Mai University, Chiangmai, Thailand.
- Sachs, R. M., Hackett, W. P., Ramina, A. and Maloof, C. 1979. Photosynthetic assimilation and nutrient diversion as controlling factors in flower initiation in *Bougainvillea* (San Diego Red) and *Nicotiana tabacum* 'Wis. 38'. In. *Photosynthesis and Plant Development* (Eds. Dr. W. Junk Pub.) The Hague, pp. 95-01.
- Sakai, W. S., Adams, C. and Braun, G. 2000. Pseudo bulb injected growth regulators as aids for year around production of Hawaiian *Dendrobium* orchid cutflowers. *Acta Hort.*, **541**:215-20.
- SAS Institute. 1990. *SAS/STAT. User's Guide*. Version 6, 4th Ed. Cary, NC, USA, **2**: 846.
- Su, W. R., Chen, W. S., Koshioka, M., Mander, L. N., Hung, L. S., Chen, W. H., Fu, Y. M., and Huang, K. L. 2001. Changes in gibberellins levels in the flowering shoot of *Phalaenopsis* hybrid under high temperature conditions when flower development is blocked. *Plant Physiol. and Biochem.*, **39**:45-50.
- Taiz, L. and Zeiger, E. 2006. *Plant Physiology*. Sinauer Assn. Inc. Pub. Massachusetts, pp. 700.